## **BioGenex**

**48810 Kato Road, Suite 100E & 200E** Fremont, CA 94538 Tel : +1 (800) 421-4149. Fax: +1 (510) 824-1490, support@biogenex.com

### DATA SHEET eFISHAML1/ETO dual color dual fusion probe

Catalog No. FP072-10X- 100µl-10 test FP072-20X- 200µl-20 test

Doc No: 932-FP072 Rev: C Date of Release: 10-Aug-2020

Material Provided: One vial of eFISH probe in hybridization buffer (RTU).

#### **Recommended detection system (Not supplied):**

Either of the following detection system is recommended depending on the automation/manual platform used:

eFISH Kit	Cat #	Description
eFISH Histo	DF-500-20XE	Automation
eFISH Cyto	DF-510-20XE	Automation

#### Intended Use:

The BioGenex eFISH AML1/ETO dual color dual fusion probe is currently available for Research use only.eFISH AML1/ETO dual color dual fusion probe is designed to be usedfor the detection of the translocationt(8;21)(q22;q22) in formalin-fixed, paraffinembeddedtissue or cells by fluorescence in situhybridization (FISH).

BioGenex eFISH AML1/ETO dual color dual fusion probe comesin hybridization buffer contains green-labeled polynucleotides(Green: excitation at 503 nm and emission at528 nm, similar to FITC), which target the AML1gene in 21q22, and orange-labeled polynucleotides(Orange: excitation at 547 nm andemission at 572 nm, similar to rhodamine), whichtarget the ETO gene in 8q22.

#### **Summary and Explanation**

Fluorescence *in situ* hybridization (FISH) is a robust technique of cytogenetic used for the detection of chromosomal aberrations, presence or absence of specific DNA sequence in native context. In this technique florescent probes bind to the target sequence of DNA in chromosome. High specificity and sensitivity coupled rapid and an accurate result has proven role of FISH in both research and diagnosis of solid tumor and hematological malignancies. As technique of cancer cytogenetics, FISH, can be used to identify genetic aberrations viz., deletions, amplification and translocation in tissue sections or within

individual cells. FISH is also used for use in genetic counseling, medicine, and species identification. FISH can also be used to detect and localize specific RNA targets in cells, circulating tumor cells, and tissue samples<sup>1,2,3,4,5</sup>.

In an FISH procedure, fixed tissue sections are pretreated to expose target DNA or mRNA sequences. An appropriately labeled probe is hybridized to the exposed target DNA or mRNA sequences in the cells. Subsequent stringent washing steps remove any probe that is non-specifically bound to the tissue section. Subsequently slides are mounted using DAPI/anti-fade and can be visualized under fluorescence microscope using appropriate filter set.

#### **Principles of the Procedure**

*In Situ* hybridization (ISH) allows the detection and localization of definitive nucleic acid sequences directly within a cell or tissue. High specificity is ensured through the action of annealing of fluorescence probe nucleic acid sequence to complementary target nucleic acid sequence. ISH techniques can be used to identify genetic aberrations like deletions, amplification, translocationin tissue sections or within individual cells.

#### **Storage and Handling**

The BioGenex eFISH AML1/ETO dual color dual fusion probe must be stored at 2-8°C protected from light and is stable through the expiry date printed on the label.

#### **Specimen Collection and Slide Preparation**

Tissues fixed in 10% (v/v) formalin are suitable for use prior to paraffin embedding and sectioning.

#### **FISH Staining procedure**

(a) The BioGenex eFISH probes are supplied in hybridization buffer and used without further dilution.

#### Protocol

Please refer to the instruction/protocol for automated or semi-automated FISH processing platform (Xmatrx<sup>®</sup>-Infinity, Xmatrx<sup>®</sup>-Nano and Xmatrx<sup>®</sup>-mini.

Further processing, such as washing and counter-staining, can becompleted according to the user's needs. For a particularly user-friendlyperformance, we recommend the use of a BioGenex eFISH Kit. These systems were also used for the confirmation of appropriateness of the BioGenex eFISH AML1/ETOdual color dual fusion probe.

**Disclaimer**: The above information is provided for reference only. Each end-user is responsible for developingand validating optimal testing conditions for use with this product.

# **BioGenex\_\_**

₩ 48810 Kato Road, Suite 100E & 200E Fremont, CA 94538 Tel : +1 (800) 421-4149, Fax: +1 (510) 824-1490, support@biogenex.com

#### Troubleshooting

Contact BioGenex Technical Service Department at **1-800-421-4149 or your local distributor** to report unusual staining.

#### **Expected Results**

The use of BioGenex eFISH AML1/ETO color dual fusion probe probein interphase and normal chromosome where there is no translocation produces two green and two orange signals separately. Where as in regions where translocation is indicated the signals appear as two green/orange fusion signals in additions to a separate single green and orange signal.

We recommend the use of atleast one control in which the t(8,21) status is known to judge the specificity of the signals.

Care should be taken not to evaluate overlapping cells, in order to avoidfalse results, e.g. an amplification of genes. Due to de-condensedchromatin, single FISH signals can appear

#### Limitations of the Procedure

Correct treatment of tissues prior to and during fixation, embedding, and sectioning is important for obtaining optimal results. Inconsistent results may be due to variations in tissue processing, as well as inherent variations in tissue. The results from *in situ* hybridization must be correlated with other laboratory findings.

#### Bibliography

- 1. Gall, J. G. and Pardue, M. L. (1969). Proc. Natl. Acad. Sci. USA63, 378 -383.
- 2. Rudkin, G. T. and Stollar, B. D. (1977). Nature 265,472-473.
- Hougaard, D. M., Hansen, H. and Larsson, L. I. (1997). Histochem. CellBiol. 108,335 -344.
- 4. Bauman, J. G., Wiegant, J., Borst, P. and van Duijn, P. (1980). *Cell Res.* 128,485 490.
- 5. O'Connor, C. (2008). Nature Education 1(1):171.
- 6. Dayyani F, et al. (2008) Blood 111: 4338-47.
- 7. Estey E and Döhner H (2006) Lancet 368: 1894-907.
- 8. Gmidène A, et al. (2011) Med Oncol 28: S5094-12.
- 9. Kievits T, et al. (1990) Cytogenet Cell Genet 53: 134-6.

- 10. Licht D (2001) Oncogene 20: 5560-79.
- 11. Vangala RK, et al. (2003) Blood 101: 270-7.
- 12. Wilkinson DG: In Situ Hybridization, A Practical Approach, Oxford University Press (1992) ISBN 0 19 963327 4.