

48810 Kato Road, Suite 100E & 200E

Fremont, CA 94538

Tel: +1 (800) 421-4149, Fax: +1 (510) 824-1490,

support@biogenex.com

DATA SHEET eFISHBCL6 Dual Color Break Apart Probe

Catalog No. FP080-10X-100µl- 10 test FP080-20X -200µl- 20 test

Doc No: 932-FP080 Rev: D Date of Release: 10-Aug-2020

Material Provided: One vial of eFISH probe in hybridization buffer (RTU).

Recommended detection system (Not supplied):

Either of the following detection systemsis recommended depending on the automation/manual platform used:

eFISH Kit	Cat #	Description
eFISH Histo	DF-500-20XE	Automation
eFISH Cyto	DF-510-20XE	Automation

Intended Use:

The BioGenex eFISH BCL6 Dual Color Break Apart Probe is currently available for Research use onlyeFISH BCL6 Dual Color Break Apart Probe is designed to be used for the detection of translocations involving the BCL6 gene at 3q27.3 in formalin-fixed, paraffinembedded tissue or cells by fluorescence in situ hybridization (FISH).

eFISH BioGenex BCL6 Dual Color Break Apart Probe in hybridization buffer. The probe contains green-labeled polynucleotides (Green: excitation at 503 nm and emission at 528 nm, similar to FITC), which target sequences mapping in 3q27.3 proximal to the BCL6 gene, and orange-labeled polynucleotides (Orange: excitation at 547 nm and emission at 572 nm, similar to rhodamine), which target sequences mapping in 3q27.3-q28 distal to the BCL6 gene.

Summary and Explanation

Fluorescence *in situ* hybridization (FISH) is a robust technique of cytogenetic used for the detection of chromosomal aberrations, presence or absence of specific DNA sequence in native context. In this technique florescent probes bind to the target sequence of DNA in chromosome. High specificity and sensitivity coupled rapid and an accurate result has proven role of FISH in both research and diagnosis of solid tumor and hematological malignancies. As technique of cancer cytogenetics, FISH, can be used to identify genetic

aberrations viz., deletions, amplification and translocation in tissue sections or within individual cells. FISH is also used for use in genetic counseling, medicine, and species identification. FISH can also be used to detect and localize specific RNA targets in cells, circulating tumor cells, and tissue samples^{1,2,3,4,5}.

In FISH procedure, fixed tissue sections are pretreated to expose target DNA or mRNA sequences. An appropriately labeled probe is hybridized to the exposed target DNA or mRNA sequences in the cells. Subsequent stringent washing steps remove any probe that is non-specifically bound to the tissue section. Subsequently slides are mounted using DAPI/antifade and can be visualized under fluorescence microscope using appropriate filter set.

Principles of the Procedure

In Situ hybridization (ISH) allows the detection and localization of definitive nucleic acid sequences directly within a cell or tissue. High specificity is ensured through the action of annealing of fluorescence probe nucleic acid sequence to complementary target nucleic acid sequence. ISH techniques can be used to identify genetic aberrations like deletions, amplification, and translocationin tissue sections or within individual cells.

Storage and Handling

The BioGenex eFISH BioGenex BCL6 Dual Color Probe must be stored at 2-8°C protected from light and is stable through the expiry date printed on the label.

Specimen Collection and Slide Preparation

Tissues fixed in 10% (v/v) formalin are suitable for use prior to paraffin embedding and sectioning.

FISH Staining procedure

- (a) The BioGenex eFISH probes are supplied in hybridization buffer and used without further dilution.
- (b) Protocol:

Please refer to the eFISH probe specific instruction/protocol for automated or semi-automated FISH processing platform (Xmatrx®-Infinity, Xmatrx®-Nano and Xmatrx®-mini.

Further processing, such as washing and counter-staining, can becompleted according to the user's needs. For a particularly user-friendlyperformance, we recommend the use of a BioGenexeFISH kit.

Disclaimer: The above information is provided for reference only. Each end-user is responsible for developingand validating optimal testing conditions for use with this



48810 Kato Road, Suite 100E & 200E

Fremont, CA 94538

Tel: +1 (800) 421-4149, Fax: +1 (510) 824-1490,

support@biogenex.com

product.

Troubleshooting

Contact BioGenex Technical Service Department at 1-800-421-4149 or your local distributor to report unusual staining.

Expected Results

The BioGenexeFISH BCL6 Dual Color Break ApartProbe is a mixture of two direct labeledprobes hybridizing to the 3q27.3-q28band. The green fluorochrome direct labeledprobe hybridizes at 3q27.3 proximalto the BCL6 gene, and the orange fluorochromedirect labeled probe hybridizes at 3q27.3-q28 distal to the BCL6 gene.

In an interphase nucleus lacking a translocationinvolving the 3q27.3 band, twoorange/green fusion signals are expectedrepresenting two normal (non-rearranged)3q27.3 loci. A signal pattern consisting of one orange/green fusion signal, one orangesignal, and a separate green signal indicate one normal 3q27.3 locus and one3q27.3 locus affected by a translocation.

Care should be taken not to evaluate overlapping cells, in order to avoidfalse results, e.g. an amplification of genes. Due to decondensedchromatin, single FISH signals can appear as small signal clusters. Thus, two or three signals of the same size, separated by a distance equal to orless than the diameter of one signal, should be counted as one signal.

Limitations of the Procedure

Correct treatment of tissues prior to and during fixation, embedding, and sectioning is important for obtaining optimal results. Inconsistent results may be due to variations in tissue processing, as well as inherent variations in tissue. The results from *in situ* hybridization must be correlated with other laboratory findings.

Bibliography

- 1. Gall, J. G. and Pardue, M. L. (1969). Proc. Natl. Acad. Sci. USA63, 378 -383.
- **2.** Rudkin, G. T. and Stollar, B. D. (1977). *Nature* 265,472-473.
- **3.** Hougaard, D. M., Hansen, H. and Larsson, L. I. (1997). *Histochem. Cell Biol.* 108,335 -344.
- **4.** Bauman, J. G., Wiegant, J., Borst, P. and van Duijn, P. (1980).. *Exp. Cell Res.* 128.485 -490.

- **5.** O'Connor, C. (2008). *Nature Education* 1(1):171.
- Joshua Weaver, Erinn Downs-Kelly, John R Goldblum, Sondra Turner,
 Sucheta Kulkarni, Raymond R Tubbs, Brian P Rubin and Marek Skacel
 .(2008).. Modern Pathology, 21, 943–949
- Hiroaki Kimura, YohDobashi, Takayuki Nojima, Hiroyuki Nakamura, Norio Yamamoto, Hiroyuki Tsuchiya, Hiroko Ikeda, Seiko Sawada-Kitamura, TakeruOyama, AkishiOoi (2013).. Int J Clin Exp Pathol 6(7):1306-1316