

48810 Kato Road, Suite 100E & 200E

Fremont, CA 94538

Tel: +1 (800) 421-4149, Fax: +1 (510) 824-1490,

support@biogenex.com

ECIREP Emergo Europe, Westervoortsedijk 60, 6827 AT Arnhem, The Netherlands

DATA SHEET eFISH PTEN/CEN 10 Dual Color Probe

Catalog No.

FP196-10XE- 100μl-10 test FP196-20XE- 200μl-20 test

Doc No: 932-FP196E Rev: A Date of Release: 21-Mar-2023

Material Provided: One vial of eFISH probe in hybridization buffer (RTU).

Recommended detection system (Not supplied):

Either of the following detection system is recommended depending on the automation/manual platform used:

eFISH Kit	Cat #	No of Tests	Description	
eFISH Histo	DF500-20XE	20	Xmatrx Automation	
eFISH Cyto	DF510-20XE	20	Xmatrx Automation	
eFISH Histo	DF520-20X	20	NanoVIP Automation	
eFISH Cyto	DF530-20X	20	NanoVIP Automation	
eFISH Histo	DF521-50X	50	NanoVIP Automation (Open system)	
eFISH Cyto	DF531-50X	50	NanoVIP Automation (Open system)	
eFISH Histo	DF522-50X	50	NanoVIP Automation (Closed system)	
eFISH Cyto	DF532-50X	50	NanoVIP Automation (Closed system)	

Intended Use:

The BioGenex eFISH PTEN/CEN 10 Dual Color Probe is currently available for **Invitro diagnostic use**. The BioGenex eFISH PTEN/CEN 10 Dual Color Probe is designed to be used for the detection of the human PTEN gene deletions as well as alpha satellites of chromosome 10 in formalin-fixed, paraffin-embedded tissue or cells by fluorescence in situ hybridization (FISH).

Summary and Explanation

BioGenex eFISH PTEN/CEN 10 Dual Color Probe comes in Formamide based hybridization buffer. The probe contains green-labeled polynucleotides (Green: excitation at 503 nm and emission at 528 nm), which target the PTEN gene, and orange-labeled polynucleotides (Orange: excitation at 547 nm and emission at 572 nm), which target sequences mapping for alpha satellite centromeric region of chromosome 10 (D10Z1)*.

Principles of the Procedure

Fluorescence *in situ* hybridization (FISH) is a robust technique of cytogenetic used for the detection of chromosomal aberrations, presence or absence of specific DNA sequence in native context. In this technique florescent probes bind to the target sequence of DNA in chromosome. High specificity and sensitivity coupled rapid and an accurate result has proven role of FISH in both research and diagnosis of solid tumor and hematological malignancies. As technique of cancer cytogenetics, FISH, can be used to identify genetic aberrations viz., deletions, amplification and translocation in tissue sections or within individual cells. FISH is also used in genetic counseling, medicine, and species identification. FISH can also be used to detect and localize specific RNA targets in cells, circulating tumor cells, and tissue samples.

In FISH procedure, fixed tissue sections are pretreated to expose target DNA or mRNA sequences. An appropriately labeled probe is hybridized to the exposed target DNA or mRNA sequences in the cells. Subsequent stringent washing steps remove any probe that is non-specifically bound to the tissue section. Subsequently slides are mounted using DAPI/antifade and can be visualized under fluorescence microscope using appropriate filter set.

Storage and Handling

The BioGenex eFISH PTEN/CEN 10 Dual Color Probe must be stored at 2-8°C protected from light and is stable through the expiry date printed on the label.

Specimen Collection and Slide Preparation

Tissues fixed in 10% (v/v) formalin are suitable for use prior to paraffin embedding and sectioning.

FISH Staining procedure

- (a) The BioGenex eFISH probes are supplied in hybridization buffer and used without further dilution.
- (b) Protocol:

Please refer to the eFISH probe specific instruction/protocol for automated or semiautomated FISH processing platform (Xmatrx®-Infinity, Xmatrx®-Nano and Xmatrx®mini.

Further processing, such as washing and counter-staining, can be completed according to the user's needs. For a particularly user-friendly performance, we recommend the use of a BioGenex eFISH kit.

These systems were also used for the confirmation of appropriateness of the BioGenex eFISH PTEN/CEN 10 Dual Color Probe.



48810 Kato Road, Suite 100E & 200E Fremont, CA 94538 Tel: +1 (800) 421-4149, Fax: +1 (510) 824-1490, support@biogenex.com

Disclaimer: The above information is provided for reference only. Each end-user is responsible for developing and validating optimal testing conditions for use with this product.

Troubleshooting

Contact BioGenex Technical Service Department at **1-800-421-4149 or your local distributor** to report unusual staining.

Expected Results

The BioGenex eFISH PTEN/CEN 10 Dual Color Probe is a mixture of an orange fluorochrome direct labeled CEN 10 probe specific for the alpha satellite sequences in chromosome 10 and a green fluorochrome direct labeled PTEN probe specific for the PTEN gene at 10q23.2-q23.31*.

In a normal interphase nucleus, two orange and two green signals are expected. In a cell with deletion of the PTEN gene locus, reduced number of green signal will be observed. Deletions affecting only parts of the PTEN gene region might result in normal signal pattern with green signals of reduced size.

However, we recommend the use of a control sample in which the 10q23* status is known to judge the specificity of the signals with each hybridization reaction.

Care should be taken not to evaluate overlapping cells, in order to avoid false results, e.g. an amplification of genes. Due to decondensed chromatin, single FISH signals can appear as small signal clusters. Thus, two or three signals of the same size, separated by a distance equal to or less than the diameter of one signal, should be counted as one signal.

Limitations of the Procedure

Correct treatment of tissues prior to and during fixation, embedding, and sectioning is important for obtaining optimal results. Inconsistent results may be due to variations in tissue processing, as well as inherent variations in tissue. The results from *in situ* hybridization must be correlated with other laboratory findings.

Bibliography

- 1. Ach T, et al. (2013) Virchows Arch 462: 65-72.
- 2. Cairns P, et al. (1997) Cancer Res 57: 4997-5000.
- 3. Dahia PLM, et al. (1999) Hum Mol Genet 8: 185-93.
- 4. Devilee P, et al. (1988) Genomics 3: 1-7.
- 5. Ettl T, et al. (2012) Br J Cancer 106: 719-26.



- 6. Ettl T, et al. (2014) Head Neck 36: 517-23.
- 7. Healy E, et al. (1998) Oncogene 16: 2213-8.
- 8. Kievits T, et al. (1990) Cytogenet Cell Genet 53: 134-6.
- 9. Li J, et al. (1997) Science 275: 1943-7.
- 10. Robertson GP, et al (1998) Proc Natl Acad Sci USA 95: 9418-23.
- 11. Swoboda A, et al. (2011) Genes Chromosomes Cancer 50: 680-8.
- 12. Weng LP, et al. (2001) Hum Mol Genet 10: 599-604.
- 13. Wilkinson DG: In Situ Hybridization, A Practical Approach, Oxford University Press (1992) ISBN 0 19 963327 4.
- 14. Yoshimoto M, et al. (2006) Cancer Genet Cytogenet 169: 128-37.
- 15. Yoshimoto M, et al. (2007) Br J Cancer 97: 678-85.

2°C 8°C	Temperature Limitation	IVD	In Vitro Diagnostic Medical Device
	Use By Date	LOT	Batch Code
NON STERILE	Non-Sterile		Consult Instructions for Use
EC REP	Representati ve in the European Community	3	BioGenex